



College of Medicine
Al-NAHRAIN UNIVERSITY

Practical Virology Lab-6- Virus Titration

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Assay of viruses Titration:

□ Biological

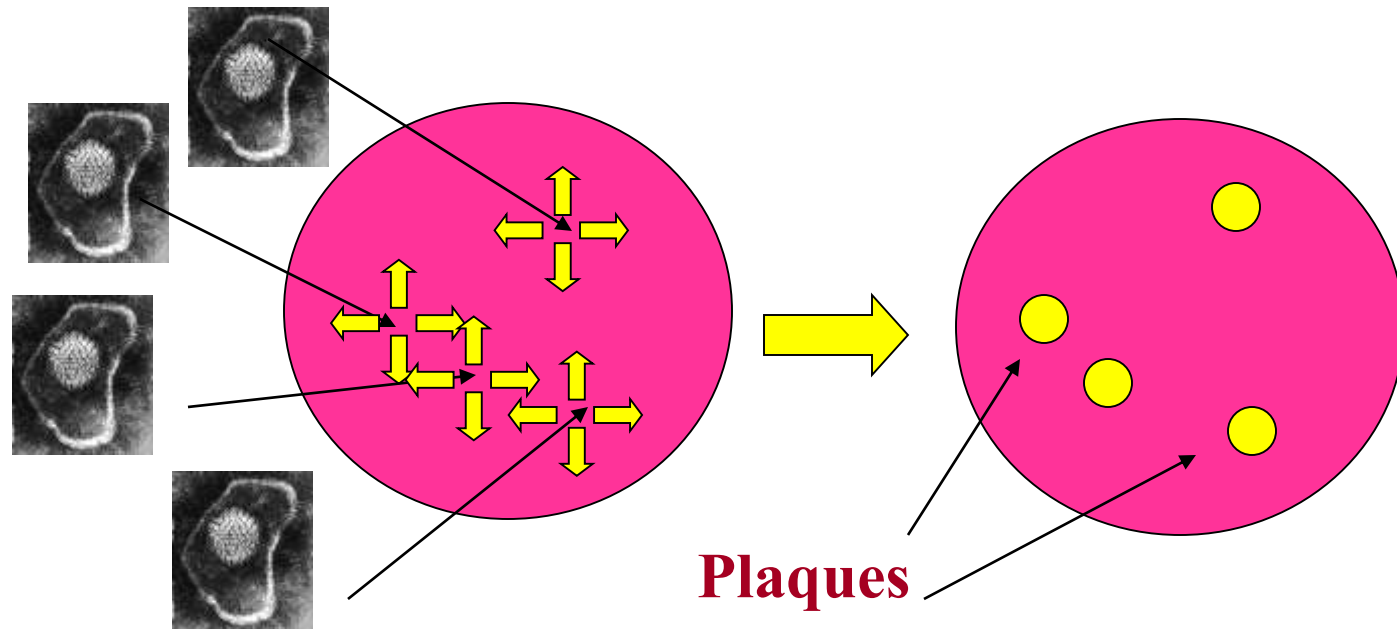
- A. Plaque assay
- B. Endpoint Method: TCID₅₀, EID₅₀, & LD₅₀.

□ Physical

- A. Hemagglutination.
- B. Direct particle count.

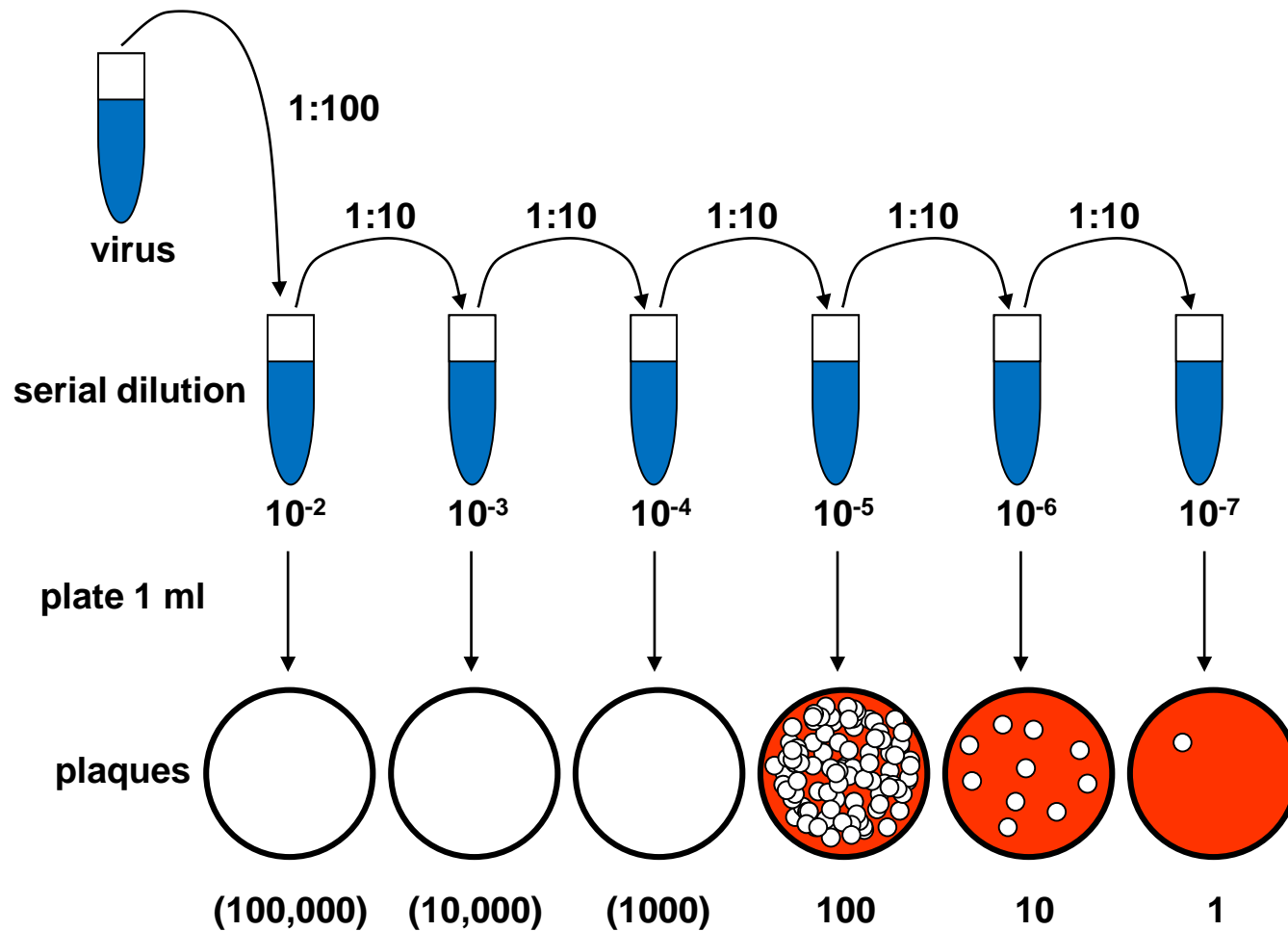
Virus Titration by Biological Methods

A- Plaque Assay



Count plaques(plaque forming unit/ml)

Procedure:

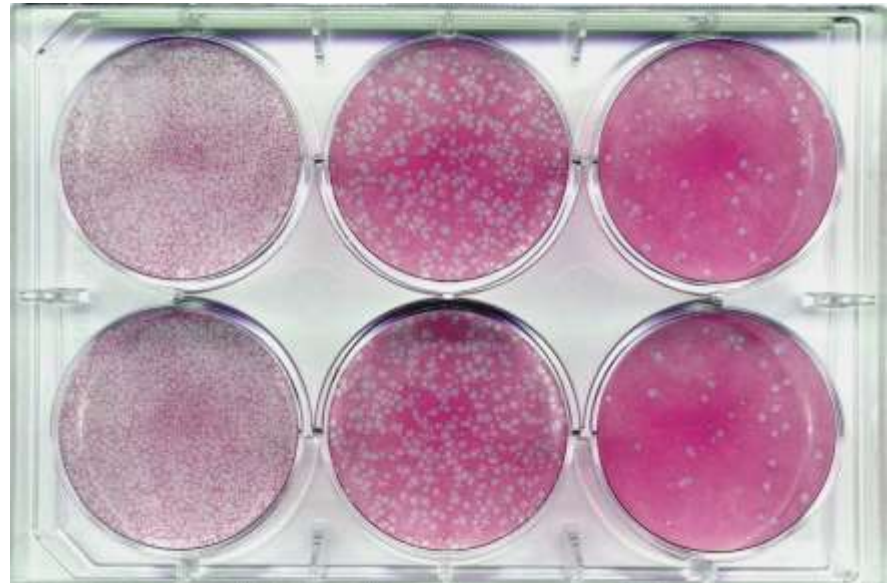


$$\text{Titer} = 1 \times 10^7 \text{ pfu/ml}$$

Plaque assay applications:

❑ Identification of the virus by very specific anti-sera.

❑ Quantitation of the virus titer in the sample (Plaque forming unit) = PFU



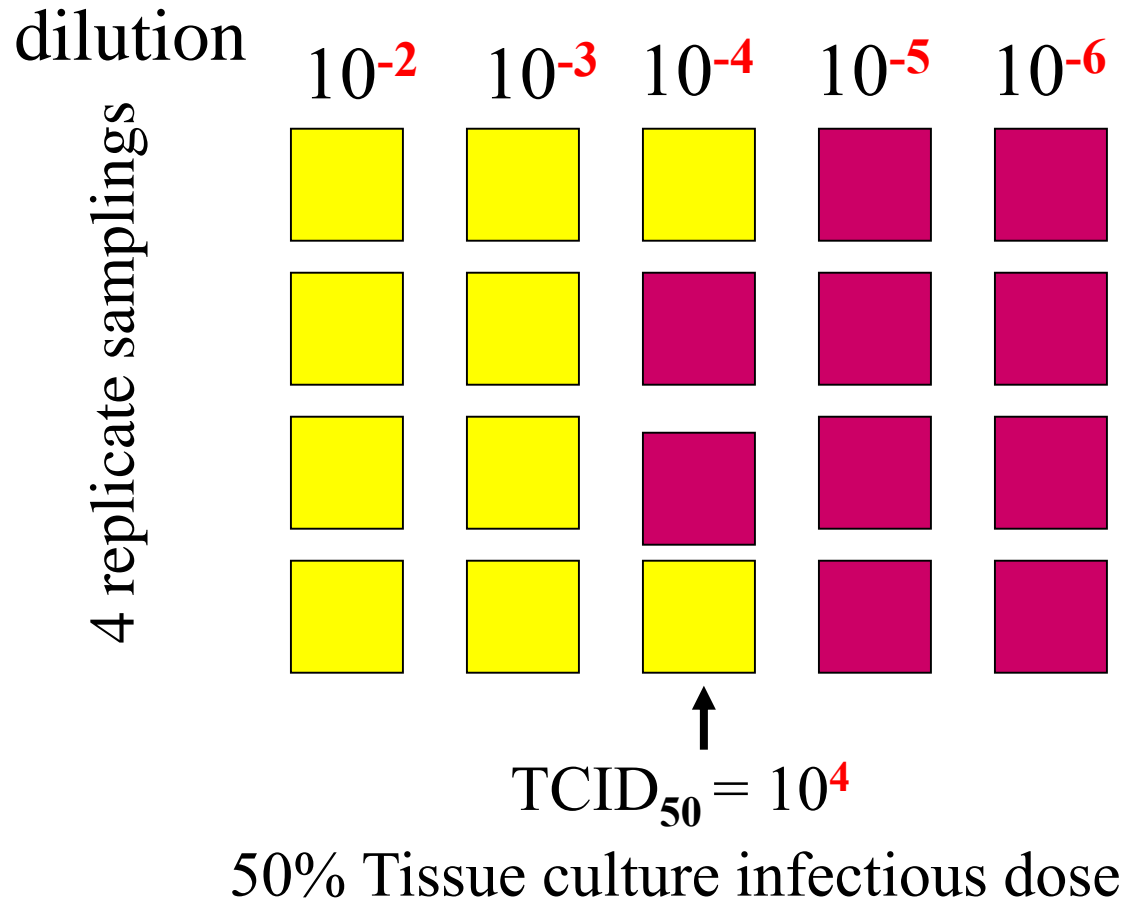
Plaque assays used to quantitate a viral stock.

Virus Titration by Biological Methods

B- TCID 50 Tissue Culture Infectious Dose 50%:

1. Prepare a monolayer of cell culture in flat bottomed wells of a microtiter plate.
2. Do serial dilution of the sample that contain the virus (e.g.10 fold dilution).
3. Inoculate each dilution in a well.
4. Incubate at 37°C & daily examine for the appearance of CPE.
5. Determine the infectivity of the virus using Karber method, (TCID 50 equals to the reciprocal of dilution that causes CPE in 50% of the cells in the monolayer in the well).

Procedure:



Endpoint Method: TCID₅₀, EID₅₀, & LD₅₀.

- **TCID₅₀**: is defined as that reciprocal of dilution of virus per unit volume required to infect & produce CPE in 50% of the cell cultures inoculated.
- **The EID₅₀**: is defined as the reciprocal of that dilution of virus per unit volume that results in death (or other observable endpoint) in 50% of the inoculated eggs.
- **The LD₅₀** : is defined as the reciprocal of that dilution of virus per unit volume that will kill 50% of the inoculated animals.

Factors that influence the LD₅₀

- The age of the animal.
- The health status of the animal.
- The strain of the animal.
- The route of inoculation.
- The strain and passage of the infecting virus.
- The size of the inoculums.
- Smaller dilution interval & the larger the number of animals per experimental group, the greater the accuracy of the results obtained.

Virus Titration by Physical Methods:

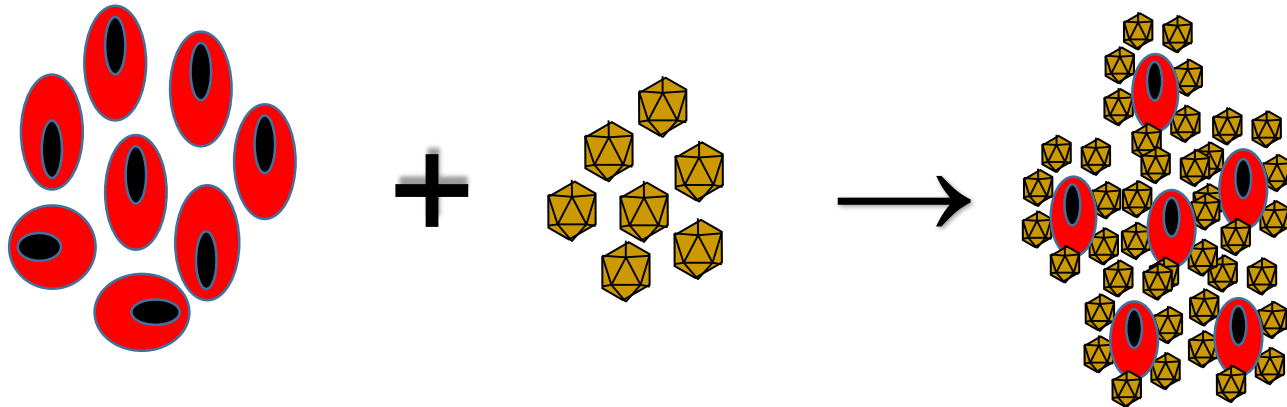
A. Hemagglutination.

- Hemagglutination (HA) test.
 - Biological phenomena.
 - Virus titration.
- Hemagglutination Inhibition (HI) test.
 - Serological test.
 - Antibody titration

B. Direct particle count.

Virus Titration by Physical Methods:

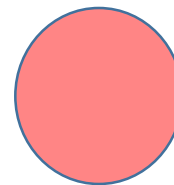
A. Haemagglutination (HA) test.



RBC suspension



HA virus



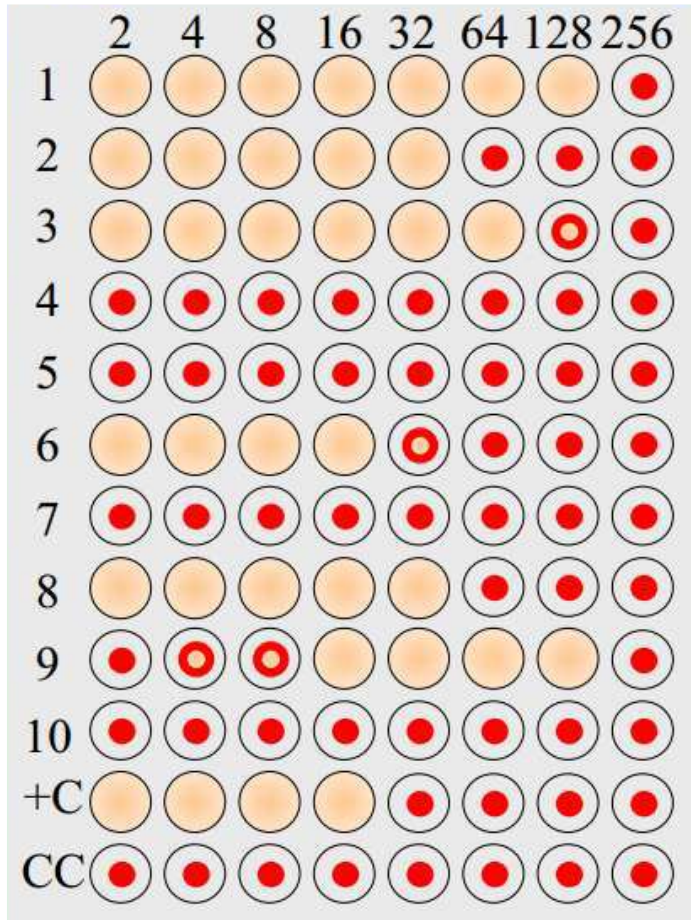
Diffuse Pattern

HA test procedure:

Materials	well No.									
(ml)	1	2	3	4	5	6	7	8	9	10
Saline	0.45	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25
Virus	0.05	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	-
Dilution	1:10	1:20	1:40	1:80	1:160	1:320	1:640	1:1280	1:2560	con
0.5%RBC	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25
incubate the plate at room temperature for 30 to 60 min.										
Check the agglutination.										

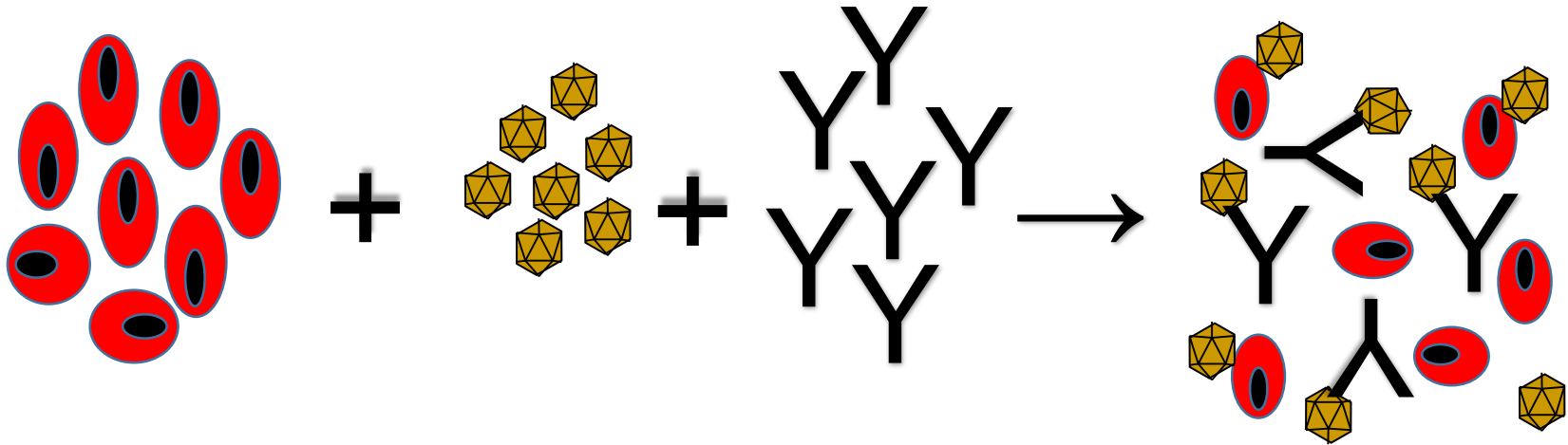
Interpretation of HA test:

HA titer = the reciprocal of highest dilution of virus giving complete HA.



Row No.	Endpoint
1	1:128
2	1:32
3	1:64
4	Neg.
5	Neg.
6	1:16
7	Neg.
8	1:32
9	1:128
10	Neg.

Haemagglutination inhibition (HI) test:



RBC suspension

HA virus

Antibody

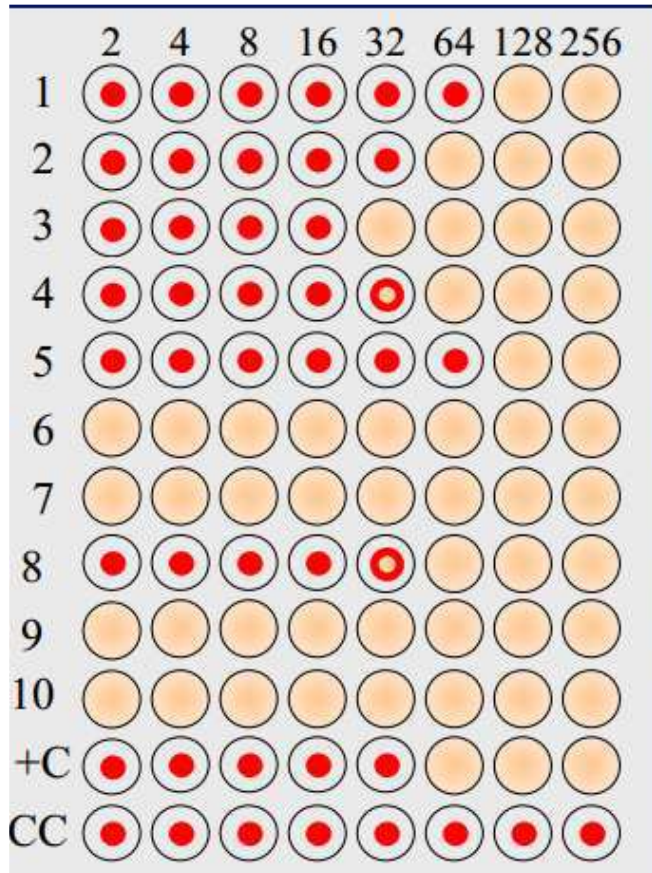
Settling Pattern

HI test procedure:

Materials	well No.									
(ml)	1	2	3	4	5	6	7	8	9	10
Saline	0.9	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25
Serum	0.1	0.25	0.25	0.25	0.25	0.25	0.25	-	0.25	-
Dilution	1:10	1:20	1:40	1:80	1:160	1:320	1:640	-	-	-
Virus	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	-	-
0.5%RBC	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	-	0.25
Mix evenly incubation at 37C for 1h										
Result	-	-	-	+	++	+++	++++	++++	-	-

Interpretation of HI test:

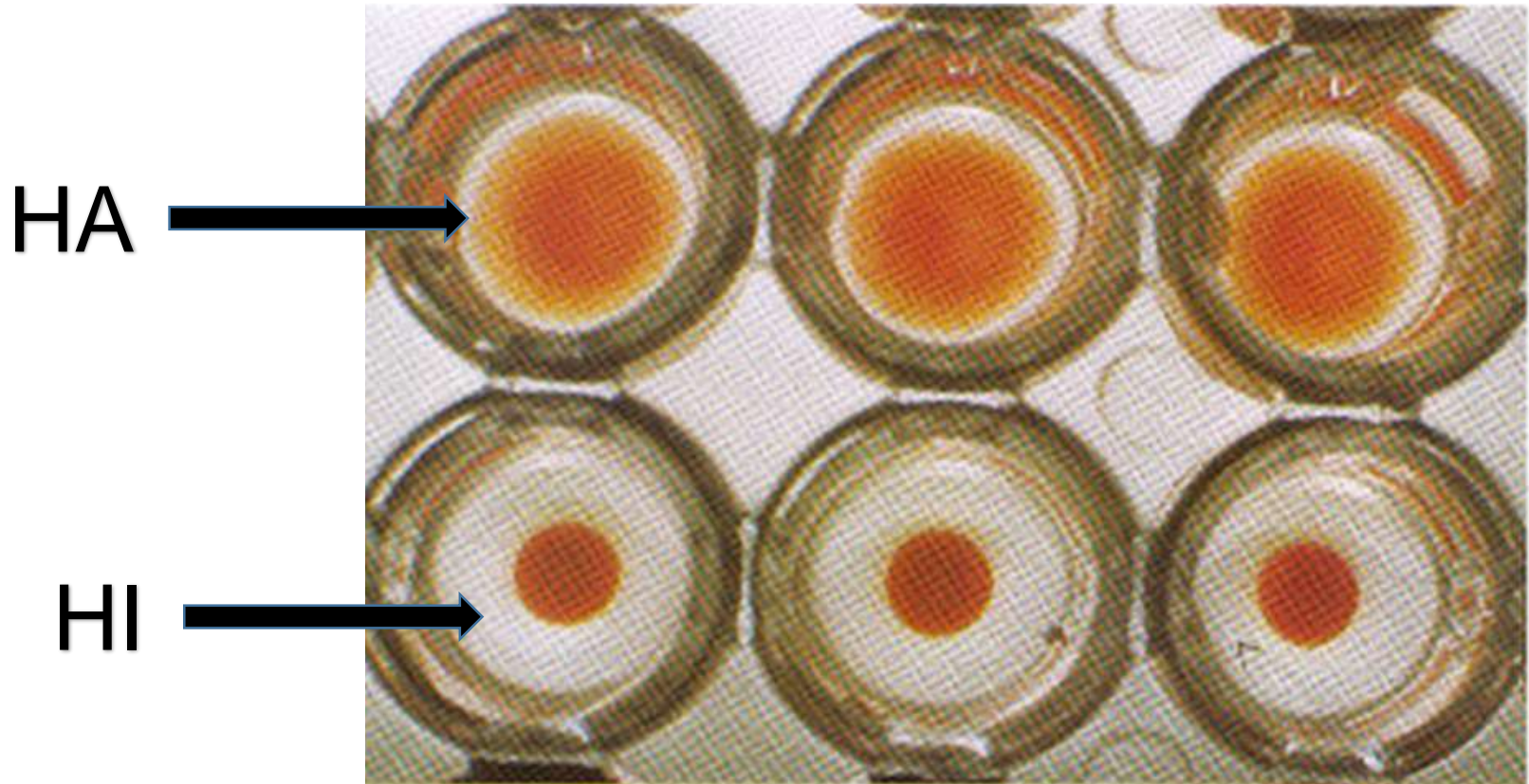
HI titer is the reciprocal of the highest dilution of the patient's serum which shows complete inhibition of agglutination.



Row No.	Endpoint
1	1:64
2	1:32
3	1:16
4	1:32
5	1:64
6	Neg.
7	Neg.
8	1:32
9	Neg.
10	Neg.

Haemagglutination Vs. Haemagglutination Inhibition

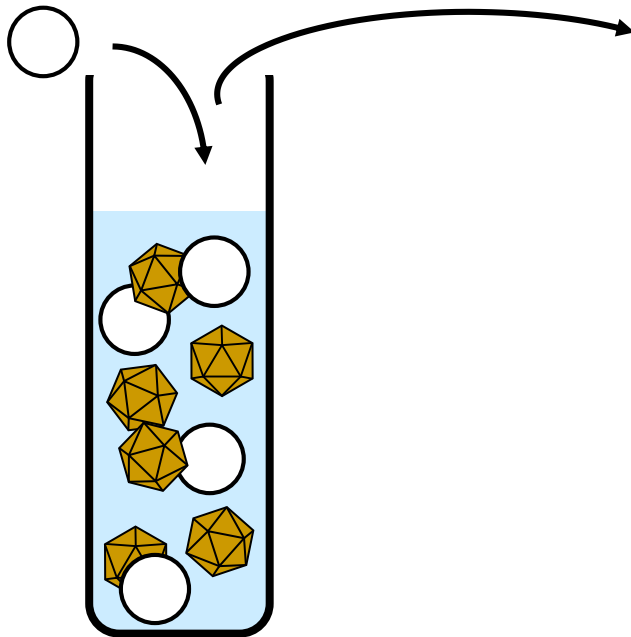
Analysis of the Results:



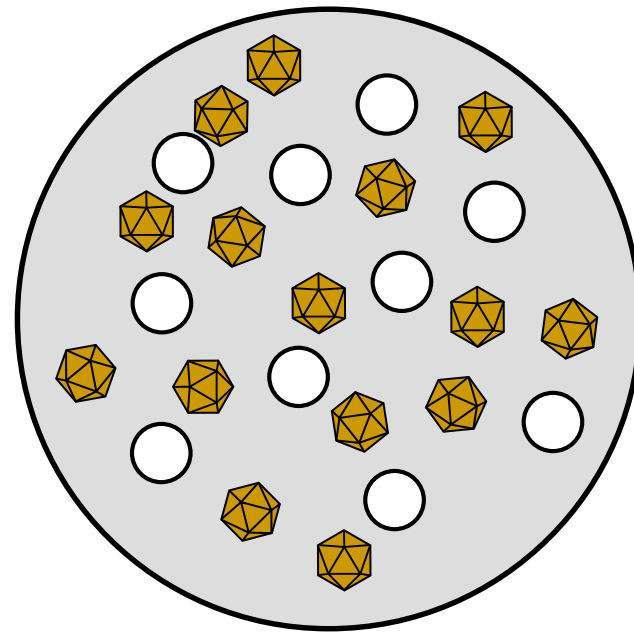
Virus Titration by Physical Methods:

B. Direct particle count.

Beads ($10^4/\text{ml}$)



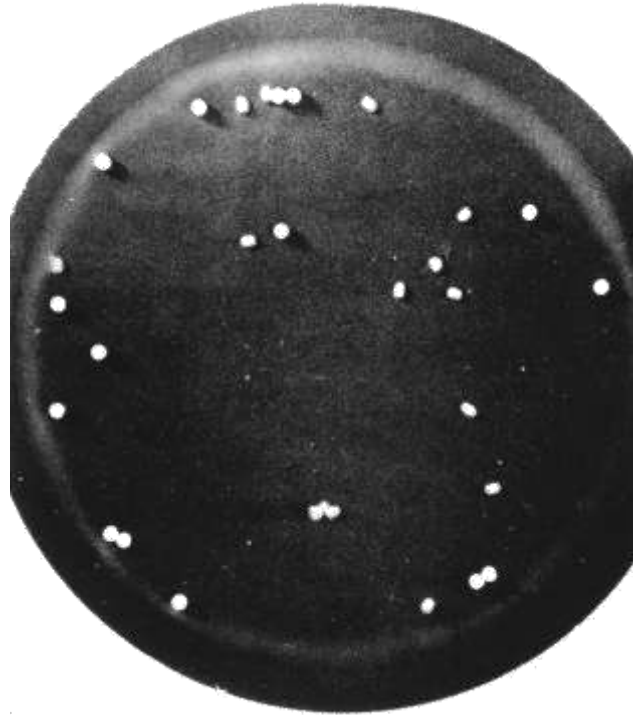
1.5×10^4 virus/ml



15 virus

10 beads

Direct particle count



Direct electron microscopic particle count. An electron micrograph of a spray droplet containing 15 **latex beads (spheres)** and 14 vaccinia **virus particles (slightly smaller, brick-shaped particles)**. (From Fields Vriology (2007) 5th edition, Knipe, DM & Howley, PM, eds, Wolters Kluwer/Lippincott Williams & Wilkins, Philadelphia Fig. 2.8)

SOME POINTS TO REMEMBER

Infectivity:

- ❑ Not every released particle is infectious.

Assays:

- ❑ Detect every particle (e.g. electron microscope)
- ❑ Detect infectious particles only (e.g. plaque assay)

THANK
YOU

